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► **To cite this version:**

Mashhour Chakouch, Gwladys Leclerc, Fabrice Charleux, Sabine Bensamoun. Phantoms mimicking the viscoelastic behavior of healthy and fibrotic livers with Magnetic Resonance Elastography technique. IEEE-ICABME 3rd International Conference on Advances In Biomedical Engineering, Sep 2015, Beyrouth, Lebanon. 10.1109/ICABME.2015.7323301 . hal-03797330

**HAL Id: hal-03797330**

**<https://hal.utc.fr/hal-03797330>**

Submitted on 4 Oct 2022

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# Phantoms mimicking the viscoelastic behavior of healthy and fibrotic livers with Magnetic Resonance Elastography technique

Mashhour K. Chakouch, *Member, IEEE*, Gwladys E. Leclerc, Fabrice Charleux, Sabine F. Bensamoun

**Abstract**—The purpose was to develop phantoms mimicking the viscoelastic properties of healthy and fibrotic livers. Three plastic phantoms composed of different concentrations (from 40% to 60%) were characterized at 60 Hz using Magnetic Resonance Elastography (MRE) tests. In parallel, the functional behavior (elasticity and viscosity) of healthy (N = 10) and fibrotic (minor: N=20 and major: N=10) livers were determined with MRE. The same MRE protocol (driver, frequency, MR sequence) was used for *in vivo* and *in vitro* experiments. MRE post-processing was realized with two methods allowing for the local measurement of the mechanical properties (shear modulus:  $\mu$ ) and the global analysis of the viscoelastic behavior through cartographies of storage ( $G'$ ) and loss ( $G''$ ) modulus. The results showed an increase of the viscoelastic ( $G'$ ,  $G''$ ) values as a function of the plastic concentration and as a function of the fibrosis level. The phantoms made of 40 %, 50 % and 60 % showed similar elastic properties of healthy (F0), minor (F1-F2) and major (F3-F4) liver fibrosis, respectively. This study has demonstrated that the developed phantoms could mimic the functional properties of healthy and fibrotic livers.

## I. INTRODUCTION

Magnetic resonance elastography (MRE) technique was applied to different healthy [1] and pathological soft tissues [2], [3] in order to provide quantitative stiffness data to the clinician from superficial to deep areas. Currently, this noninvasive MRE method is only used in clinical practice for liver test, allowing the assessment of the fibrosis level [4]. This diagnosis was based on the stiffness (shear modulus) measurement revealing higher values for pathological tissues [5].

Subsequently, MRE technique was extensively improved for the characterization of the viscoelastic behavior, revealing the fluid and solid components of the soft tissues [6]. Thus, the viscoelastic properties of the liver [7], brain [6] and muscle [8] were determined with multi-frequency MRE tests. These measurements were obtained with the

development of imaging sequences and reconstruction methods [9] in order to acquire the cartographies of the storage modulus ( $G'$ ) and the loss modulus ( $G''$ ) which compose the viscoelastic properties.

All of these developments were mainly tested on phantoms composed of different materials (wirosil [10], agarose [11], bovine gel [12]) mimicking the elastic properties of soft tissues [13] and dynamic organs [10]. Indeed, at first organic gels (agarose, bovine) were used for their accessibilities but the main inconvenience was the stability of the material as a function of time. Subsequently, wirosil phantoms were used due to their stability compared to the organic gels. However, this material showed high stiffness which was not adapted for liver tissue. Some phantoms (CIRS Inc) were commercially available for ultrasound elastography [14], but the inconvenience was the fixed Young's modulus, determined only at low frequencies, provided by the constructor. In the literature many studies created their own phantoms composed of polymer (PVC) [15] in order to control and perform different ranges of stiffness corresponding to biological tissues and to produce stable elastic properties as a function of time [16]. Recently, Nguyen's team has developed oil-in-gelatin phantoms to measure the shear modulus and the viscosity [17].

The phantoms are considered as a benchmark for medical imaging techniques. Thus, the purpose of this study was to develop phantoms mimicking the viscoelastic properties of different healthy and fibrotic livers using MRE technique.

## II. MATERIALS AND METHODS

### A. Phantom preparation

Homogeneous phantoms were created with different concentrations of liquid plastic (Plastileurre, Bricoleurre, France) and softener (Assouplissant, Bricoleurre, France) in order to mimic the different stages of the liver fibrosis. The liquid plastic, also called plastisol, is a suspension of PVC particles in a solvent. The mixture was adjusted from 40% to 60% with a step concentration of 10%, in order to progressively increase the stiffness of the media, and heated to 177°C. Subsequently, the solution was poured into aluminum molds, which were resistant to high temperature, in order to create three homogeneous parallelepiped (length: 14 cm, width: 7 cm, and height: 5 cm, Fig. 1) phantoms used for MRE technique. The different molds were left to cool to room temperature (23°C) until the phantoms solidified. The density of the phantoms was experimentally evaluated

\*Research supported by Sorbonne University, Université de Technologie de Compiègne.

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( $1000 \pm 47 \text{ kg/m}^3$ ). Then, the phantoms were preserved and stocked at room temperature ( $23^\circ\text{C}$ ).

### B. Phantom Magnetic Resonance Elastography (MRE) tests

The phantom was placed inside a head coil in a 1.5T MRI machine (Signa HDx, GE) (Fig. 1). A cylindrical driver, currently used in clinical practice for liver MRE test, was positioned below the phantom. Moreover, a soft MRI cushion was inserted between the phantom and the driver in order to avoid motion artifacts during the test. The driver is connected via a long tube to a loudspeaker, located in the MRI technical room, which generates air pressure at low frequency 60 Hz. The phase images, showing the displacement of the shear waves inside the parallelepiped phantoms, were collected with: four offsets, a gradient echo sequence, an acquisition matrix of  $256 \times 64$  (interpolated to  $256 \times 256$ ), a flip angle of  $45^\circ$ , a field of view of 20 cm, TR equal to 50 ms and TE corresponding to the minimum echo time allowing for motion encoding.

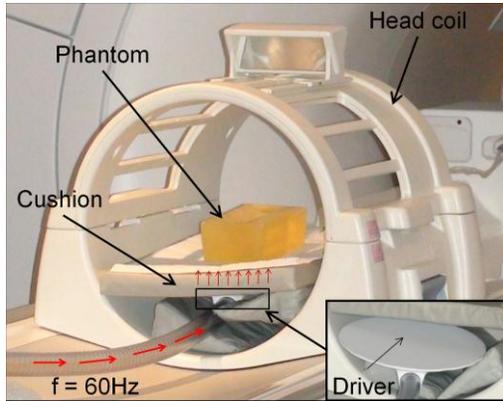


Figure 1. Set up of the magnetic resonance elastography tests performed on the different phantoms

### C. Liver Magnetic Resonance Elastography (MRE) tests

Healthy participants ( $N = 10$ ), with no history of liver disease, were recruited and two groups of patients characterized with minor ( $N = 20$ ) and major ( $N = 10$ ) fibrosis were recruited from the alcoholism department. The stage of fibrosis was determined with the Fibroscan (EchoSens) exam due to the risk induced by the biopsy which is an unnecessary procedure for alcoholic patients. The classification of the level of fibrosis was based on the METAVIR Score [18] with F0: healthy liver, F1-F2: minor fibrosis and F3-F4: major fibrosis.

Subject were placed in a supine position in a 1.5T MRI machine (Signa HDx, GE), and the cylindrical liver acoustic driver, used for the previous phantom MRE tests (Fig. 1), was positioned in contact with the ribcage at the same level as the diaphragm. Similarly to the phantom, shear waves were propagated within the liver at a single frequency of 60 Hz. Phase images (4 offsets) were recorded with a motion sensitizing gradient echo sequence, a flip angle of  $30^\circ$ , a field of view between 36 and 48 cm, a  $256 \times 64$  acquisition matrix, a TE = 26.8 ms and a TR = 100 msec. The total scan time was 32 seconds, corresponding to two breath-holding periods of 16 seconds.

### D. MRE Post-processing

The recorded phase images underwent post-processing by applying a mask, which removed the noise located in the background of the image. The viscoelastic properties of the phantoms and liver tissues were obtained with the two following methods. The first one provides a local stiffness measurement by prescribing a 1D profile drawn along the direction of the shear wave propagation (Fig.2, 3).

Assuming that the media (phantom or liver tissue) was linearly elastic, isotropic, incompressible and homogeneous, the local shear stiffness ( $\mu$ ) representing the local elasticity was calculated using the following equation  $\mu = \rho \cdot (f \cdot \lambda)^2$ , where  $\rho$  is the density assumed to be closed to the water ( $1000 \text{ kg/m}^3$ ). The wavelength ( $\lambda$ ) was determined by considering the distance between consecutive picks. The second method applied an inversion algorithm to the phase images to represent the viscoelastic properties [19] through the cartographies of the storage ( $G'$ ) and the loss ( $G''$ ) modulus. Regions of interests (ROI) were placed on cartographies to measure the average data with the standard deviation.

## II. RESULTS

### A. Viscoelastic properties of the phantoms as a function of the plastic concentration

The results of the MRE tests performed on the phantoms are summarized in Fig. 2. The phase images revealed a clear propagation of the shear waves within the different phantoms with an increase of the wavelength as a function of the plastic concentration. The results of the local ( $\mu$ ) and global ( $G'$ ) elastic values were in the same range and increase with the level of the plastic concentration.

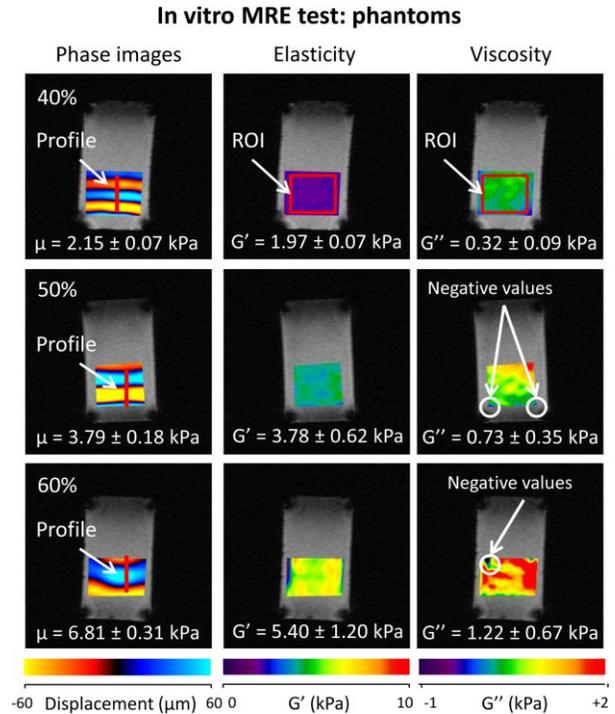


Figure 2. Phase images (60 Hz) with a red profile providing the local stiffness for the different phantoms. Cartographies of  $G'$  and  $G''$ , obtained with inversion algorithm, provide a global stiffness measured inside the represented ROI

The range of viscous data was smaller (from 0.24 kPa to 1.22 kPa) compared to the range of the  $G'$  values (from 0.89 kPa to 5.40 kPa). The viscous properties increase also as a function of the plastic concentration (Fig. 2).

### B. Viscoelastic properties of the healthy and fibrotic livers

Phase images showed a clear propagation of the shear waves within the healthy and fibrotic livers, and as expected the wavelength increased with the level of fibrosis (Fig. 3). The average storage modulus ( $G'$ ) for healthy, minor and major fibrosis were  $1.82 \pm 0.23$  kPa,  $3.78 \pm 0.74$  kPa and  $5.52 \pm 1.03$  kPa, respectively. The loss modulus ( $G''$ ) was in the same range for healthy ( $0.51 \pm 0.17$  kPa) and minor ( $0.56 \pm 0.25$  kPa) fibrosis while a slight increase was measured for the major ( $0.84 \pm 0.44$  kPa) fibrosis.

#### In vivo MRE test: liver tissues

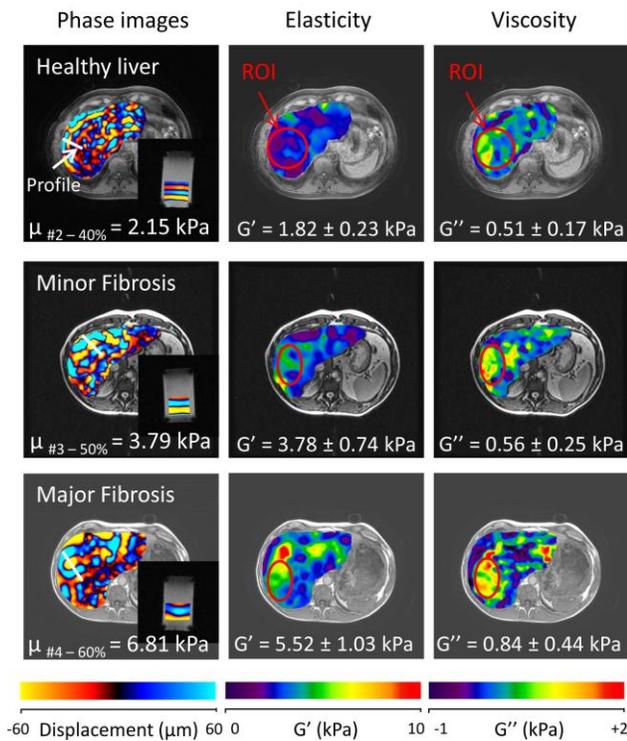


Figure 3. Phase images and cartographies ( $G'$  and  $G''$ ) obtained for healthy and fibrotic liver.

### D. Analogy between the MRE viscoelastic properties of the phantoms and different biological tissues (livers, ...)

MRE technique was also applied, by the same team, with the same set up (60 Hz, liver driver) to other tissues (spleen, kidney and muscle).

The result of the elastic properties of the phantom 40% ( $G'_{40\%} = 1.97 \pm 0.07$  kPa) mimics the elastic behavior of the F0 liver ( $G'_{F0} = 1.82 \pm 0.23$  kPa). Furthermore, the elastic behaviors of the psoas ( $2.60 \pm 0.25$  kPa) [20] and vastus medialis ( $2.64 \pm 0.20$  kPa) [8] in passive (i.e. at rest) condition were also in the same range as the phantoms made with 40 % of plastic.

The phantom with 50 % of plastic revealed elastic property ( $G'_{50\%} = 3.78 \pm 0.62$  kPa) close to minor (F1,

F2) fibrosis stages ( $G'_{\text{minor}} = 3.78 \pm 0.74$  kPa). This phantom could be used to mimic the elastic properties of these liver stages. In comparison to other tissues such as the kidney ( $4.37 \pm 0.59$  kPa) [20], the spleen ( $4.75 \pm 0.70$  kPa) [20] and the sartorius muscle ( $4.13 \pm 0.69$  kPa) [8] at rest, the phantom 50% could be used to represent the elastic properties of these tissues.

The last phantom (60 %) had elastic properties ( $G'_{60\%} = 5.40 \pm 1.20$  kPa) corresponding to the ones of the major liver fibrosis (i.e. stage F3-F4) ( $G'_{\text{major}} = 5.52 \pm 1.03$  kPa). In comparison to muscle studies, this phantom could be used to mimic the elastic properties of the semitendinosus (ST) muscle at rest ( $\mu_{\text{ST}} = 5.32 \pm 0.10$  kPa) [1].

Concerning the viscous properties, only the 50 % phantom revealed data ( $G''_{50\%} = 0.73 \pm 0.35$  kPa) close to the major fibrosis ( $G''_{\text{major}} = 0.84 \pm 0.44$  kPa).

## III. DISCUSSION

Magnetic resonance elastography (MRE) is now a clinical liver test to diagnose the extent of liver fibrosis. The further development of MRE methods has revealed utility for other medical applications (brain, breast). It is well known that the phantoms have always been considered as a necessary object for the calibration of imaging sequences used by CT scanners, MRI machines or ultrasound (US). To our knowledge, any phantom dedicated to MRE technique was developed by imaging companies while ultrasound elastography employs specific commercial phantoms for the characterization of breast and liver tissues. These tests objects are certified by the company as compatible with MRI machines, and it was therefore expected to get MRE phase images by applying MRE sequence. However, the recent liver phantom (Model 057, CIRS, Norfolk, Virginia, USA), used for US elastography, was not MRE compatible and it was impossible to record phase images using the MRE liver sequence. As a consequence, there is a real clinical need for a MRE benchmark before performing in vivo MRE tests. The originality of the present study was to develop plastic phantoms allowing for the simulation of the viscoelastic properties of liver tissues enabling clinicians to choose the appropriate phantom as a function of the fibrosis levels.

In addition to the simulation of the functional (elasticity, viscosity, density) behavior of the biological tissue, the development of a phantom also required to take into account the characteristics (driver, frequency, MR sequence) of the MRE protocol used in clinic. Indeed, the material properties (pneumatic, mechanical) and the shape (round, tube) of the driver were key points to ensure the generation of the shear wave within the tissue [15]. Thus, in the present study the same driver as the one applied for liver MRE tests in clinical practice was used in order to attenuate the eventual effects of the driver on the shear wave propagation. Furthermore, the frequency applied during the MRE test is also an important parameter of the protocol due to the frequency-dependence, or viscoelastic properties of the soft tissues. In the literature, the current range of frequency performed on human tissues, with MRE technique, is between 25 Hz and 110 Hz [6], [8]. Therefore, the developed phantoms should work for this

biological range of frequencies. Phantoms made of wirosil were extensively used by other studies [10], to obtain a clear propagation of the shear wave within the material, but the mechanical behaviors (144 kPa) were not comparable to biological soft tissues. Indeed, the use of wirosil phantoms required higher MRE frequencies (200 Hz) [10] compared to soft tissues (for instance: 60 Hz for the liver, 90 Hz for the muscle). Subsequently, plastisol material was used, enabling one to vary the plastic liquid concentration, in order to better mimic the elastic properties of biological tissues and to use similar frequency MRE ranges as in vivo studies. Thus, Baghani et al. (2009) have shown an increase of shear stiffness with the level of concentration using ultrasound elastography [21].

The development of the present phantoms revealed some shape limitations related to the chosen dimension. Indeed, the height was not appropriate for the large wavelength obtained for high plastic concentration. Therefore, it was difficult to visualize at least one wavelength for the stiffer phantom (60 %), and higher molds must be used for future MRE tests. Moreover, the inversion algorithm used in this present study should be improved due to the presence of negative viscous values. These data were found in the boundary areas of the loss modulus cartography due to the reflected waves and the presence of noise. This phenomenon was emphasized for a stiffer media. Thus, these zones were carefully avoiding during the selection of the region of interest (ROI).

This preliminary set of phantoms revealed the possibility to mimic the elastic (storage modulus) properties of healthy and pathological livers. In perspective, the phantom must be further developed to better characterize the viscous properties of the livers. In addition, the composition of the phantoms may be enhanced to have a more real phantom to mimic the anisotropic and inhomogeneous behavior of the brain, skeletal muscle, kidney, etc ...

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