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## Research Article



# Biocompatibility study of lithium disilicate and zirconium oxide ceramics for esthetic dental abutments

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## ABSTRACT

**Purpose:** The increasing demand for esthetically pleasing results has contributed to the use of ceramics for dental implant abutments. The aim of this study was to compare the biological response of epithelial tissue cultivated on lithium disilicate (LS<sub>2</sub>) and zirconium oxide (ZrO<sub>2</sub>) ceramics. Understanding the relevant physicochemical and mechanical properties of these ceramics will help identify the optimal material for facilitating gingival wound closure.

**Methods:** Both biomaterials were prepared with 2 different surface treatments: raw and polished. Their physicochemical characteristics were analyzed by contact angle measurements, scanning white-light interferometry, and scanning electron microscopy. An organotypic culture was then performed using a chicken epithelium model to simulate peri-implant soft tissue. We measured the contact angle, hydrophobicity, and roughness of the materials as well as the tissue behavior at their surfaces (cell migration and cell adhesion).

**Results:** The best cell migration was observed on ZrO<sub>2</sub> ceramic. Cell adhesion was also drastically lower on the polished ZrO<sub>2</sub> ceramic than on both the raw and polished LS<sub>2</sub>. Evaluating various surface topographies of LS<sub>2</sub> showed that increasing surface roughness improved cell adhesion, leading to an increase of up to 13%.

**Conclusions:** Our results demonstrate that a biomaterial, here LS<sub>2</sub>, can be modified using simple surface changes in order to finely modulate soft tissue adhesion. Strong adhesion at the abutment associated with weak migration assists in gingival wound healing. On the same material, polishing can reduce cell adhesion without drastically modifying cell migration. A comparison of LS<sub>2</sub> and ZrO<sub>2</sub> ceramic showed that LS<sub>2</sub> was more conducive to creating varying tissue reactions. Our results can help dental surgeons to choose, especially for esthetic implant abutments, the most appropriate biomaterial as well as the most appropriate surface treatment to use in accordance with specific clinical dental applications.

**Keywords:** Ceramics; Dental abutments; Dental esthetics; Embryo culture techniques

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**Conflict of Interest**

No potential conflict of interest relevant to this article was reported.

**INTRODUCTION**

Ceramics used in medical fields have evolved rapidly over the last 20 years. In particular, dental glass ceramics have been developed because they have advantageous physicochemical and cosmetic properties [1]. Ceramics therefore may replace metallic materials in the oral cavity, maintaining the same high quality level in dental rehabilitation [2].

Introduced to dental markets over a decade ago, all-ceramic systems covered a wide range of indications, from thin veneers used in single-tooth restorations to crowns and wide-span bridges [3]. Titanium abutments were a standard option for dental implant-supported restorations. Due to potential recession of the soft tissue margin in oral implants [4] and coloration of the gingiva [5], the all-ceramic system helps maintain the esthetics of the implant, as demonstrated by the widespread current use of this system [6]. Ceramic abutments, fabricated from yttrium stabilized-zirconium oxide (ZrO<sub>2</sub>), have been developed for their color, which is similar to that of teeth, high loading strength, tissue tolerability, and intrasulcular design enhancement [7]. Transformation toughening of ZrO<sub>2</sub> results in extremely high component stability and extraordinary bending and tensile strength, as well as fracture and chemical resistance [8,9]. These properties allow ZrO<sub>2</sub> to self-repair micro-crack initiation by stopping crack propagation. ZrO<sub>2</sub> is considered to be a highly biocompatible material, and has already been used in medical applications such as artificial hip joints.

As a result of patient demand, veneers and crowns are currently available in ZrO<sub>2</sub> or, recently, in lithium disilicate (LS<sub>2</sub>) ceramic [1,2]; they can be used with the press technique as well as computer-assisted design and manufacturing technology [10]. In this way, dental zirconia has been used daily by dental practitioners (e.g., IPS e.max<sup>®</sup> ZirCAD, Ivoclar Vivadent SAS, Saint-Jorioz, France) which is an yttrium-stabilized ZrO<sub>2</sub>. Its biological properties are very satisfactory and clinical results have proven its suitability for dental applications. Featuring a flexural strength of 900 MPa, more than double that of glass ceramic (e.g., IPS e.max<sup>®</sup> Press or CAD, Ivoclar Vivadent SAS), this biomaterial can be used for almost all applications for which metal has exclusively been used up to now, especially for the posterior teeth, long bridges, and implant armatures [8]. Nevertheless, dental zirconia ceramics are considered less esthetic than glass ceramics due to their white shade and high opacity. This is the fundamental problem for dental implant abutments; however, LS<sub>2</sub> ceramic has a similar appearance to natural teeth and could therefore improve the esthetic outcomes. The esthetic characterization of implant abutments and its ceramic restorations involves the reflection and transmission of light. IPS e.max<sup>®</sup> LS<sub>2</sub> is a high-strength ceramic material with 360–400 MPa of flexural strength. Moreover, this biomaterial was recently reported to be one of the most robust and durable all-ceramic systems tested to date [8].

In order to be a viable and superior treatment choice, ceramic restorations must be cosmetically and functionally appropriate. The surface and biological requirements in dental implantology are as follow: (1) no cell adhesion or proliferation on the esthetic crown for easy cleaning; (2) cell adhesion and proliferation around the esthetic abutment for a tight junction; and (3) strong cell adhesion under the epithelia-conjunctive junction to preserve the dental implant from buccal bacteria for an extended period.

The aim of this *in vitro* study was to compare 2 polished biomaterials for cytocompatibility and biological response. We used a combination of contact angle assessments and

interferometry measurements to determine surface wettability. An organotypic culture of chicken epithelium was then performed.

## MATERIALS AND METHODS

Two types of biomaterials were used (IPS e.max<sup>®</sup> Press, IPS e.max<sup>®</sup> ZirCAD, Ivoclar Vivadent SAS). The first was an LS<sub>2</sub> ceramic (IPS e.max<sup>®</sup> Press, Ivoclar Vivadent SAS) composed of silicon dioxide (SiO<sub>2</sub>) (57%–80%), lithium oxide (Li<sub>2</sub>O) (11%–19%), potassium superoxide (KO<sub>2</sub>) (0%–13%), phosphorus pentoxide (P<sub>2</sub>O<sub>5</sub>) (0%–11%), ZrO<sub>2</sub> (0%–8%), zinc oxide (ZnO) (0%–8%), and other oxides and pigments (0%–10%). The second one was a dental yttrium-stabilized ZrO<sub>2</sub> (IPS e.max<sup>®</sup> ZirCAD, Ivoclar Vivadent SAS) composed of ZrO<sub>2</sub> (87%–95%), hafnium oxide (HfO<sub>2</sub>) (1%–5%), aluminium oxide (Al<sub>2</sub>O<sub>3</sub>) (0%–1%), yttrium oxide (Y<sub>2</sub>O<sub>3</sub>) (4%–6%), and other oxides and pigments (0%–8%). The control samples used Thermanox<sup>®</sup> (Thx), a cell culture-treated plastic (Nunc<sup>®</sup> batch #628934) showing excellent cell migration properties associated with low cell adhesion. Thx is a very smooth material that is traditionally used for cell culture control samples.

### Sample preparation

#### LS<sub>2</sub> ceramic samples

Square samples (1.4×1.4×0.1 cm) were separately obtained by the loose-wax technique and completely fired. All samples were cleaned by an acid (Invex<sup>®</sup>, Ivoclar Vivadent SAS) and sanded in house with glass-bead blasting before use (raw-LD). Then, a surface treatment was performed using manual polishing. The mechanical polishing was done by the same operator with the Optrafine<sup>®</sup> pack (batch #NL1757, Ivoclar Vivadent SAS). A manual dental piece was used without water spray: 15 seconds with a large drill (DC 83103040, Komet, Paris, France) at 30,000 rpm, 15 seconds with a dark-blue diamond polisher in cup shape at 7,000 rpm (polisher P), 15 seconds with a light-blue diamond finisher in cup shape at 7,000 rpm (finisher F), and 15 seconds with a small brush accompanied by polishing paste (Optrafine<sup>®</sup> HP, lot JL1606, Ivoclar Vivadent SAS) at 7,000 rpm (pol-LD).

Eighty square samples were set up: 40 samples without a surface treatment (raw-LD) and 40 samples with polishing (pol-LD).

#### ZrO<sub>2</sub> ceramic samples

The zirconia blocks (size MOO B85 L-22; batch #P01949; enlargement factor, 1.228) were cut into plates (1.5×1.5×0.2 cm) using a rotating diamond saw before sintering. The plates were then ground on the surface using silicon carbide particles and water. The samples were dried afterward at 80°C–120°C for 2 hours using a drying furnace. They were then fired in a Programat S1 at 1,500°C (IPS e.max<sup>®</sup> ZirCAD program, Ivoclar Vivadent SAS) with a holding time of 30 minutes at the maximum temperature. The samples were completely sintered before use (raw-Zr). In order to achieve a reasonable polishing for flat surfaces using dental instruments, polishing was carried out after the final densification in the S1 using a Struers TegraPol-35 grinding/polishing unit (300 rpm). After sintering, apex diamond grinding discs were applied (20-, 6-, and 0.5-μm diamond wheels). Eighty square samples were also prepared, including 40 raw samples (raw-Zr) and 40 polished samples (pol-Zr).

## Physicochemical surface characterizations

### Contact angle measurements

Contact angles of water droplets were determined at ambient temperature (21°C–24°C) with a drop shape analysis apparatus (DSA-10, Krüss GmbH, Hamburg, Germany). The measuring system recorded the drop shape by a charge-coupled device (CCD) camera and determined the contact angles using image analysis software, taking into account the entire drop shape [11].

Six water droplets were deposited on each surface, and the values of the contact angle (°) represent the averages of 12 contact angle measurements (6 on the right side and 6 on the left side).

### Scanning white-light interferometry

A Zygo® NewView 200 (Zygo Corporation, Middlefield, CT, USA) apparatus was used with frequency domain analysis to generate quantitative 3-dimensional images of surfaces [12]; measurements were made using a white-light filter based on a center wavelength of 600 nm, with a bandwidth of 125 nm. The interference patterns were recorded by a CCD camera, and each measurement contained 320×240 data points. Three objectives with magnifications of ×2.5, ×10, and ×50 were used, and 3 images were recorded for each magnification. The scan size and sampling interval were fixed by the magnification of the optical system. For the magnification of ×2.5, the sampling interval was 8.8 μm in both directions. The vertical resolution was lower than 1 nm, while lateral resolution of the microscope was limited by the aperture of the objective. This technique was used to determine the biomaterial surface roughness.

### Scanning electron microscopy

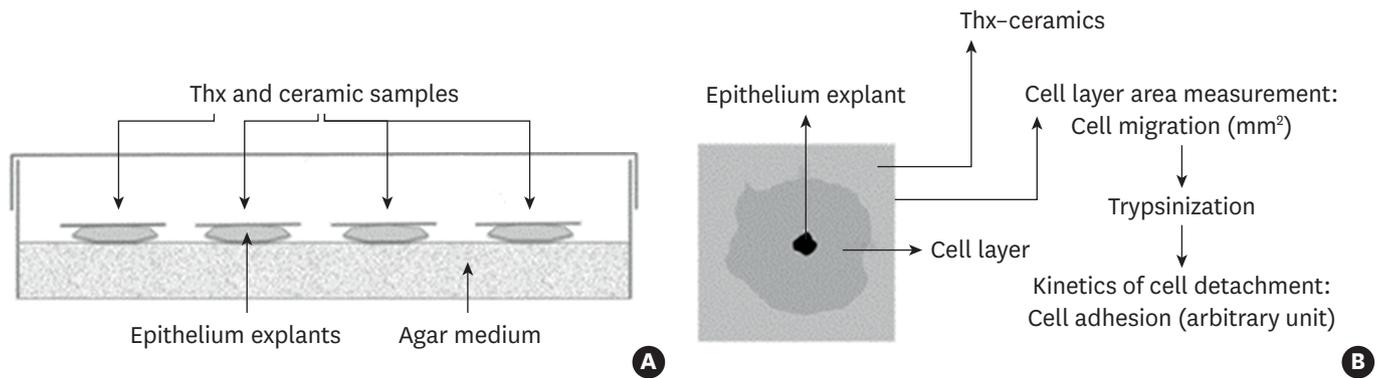
Samples of tissues cultivated on the different materials were rinsed in phosphate-buffered saline (PBS), fixed in 3% glutaraldehyde in Rembaum buffer (pH 7.4) for 1 hour [13], dehydrated in a series of graded alcohols, critical-point dried using CO<sub>2</sub> (Polaron Instrument Inc., Nottingham, UK), sputter-coated with gold (Polaron Instrument Inc.) and examined with a Philips scanning electron microscope (ESEM FEG XL 30, Philips, Amsterdam, The Netherlands).

## Organotypic culture method

Before culture, the ceramic samples were ultrasonically cleaned in acetone for 10 minutes, then rinsed in distilled water (10 minutes), and autoclaved at 120°C for 20 minutes.

An organotypic culture method [14] was used to examine the cellular response to ceramic samples (Figure 1A). Culture dishes were pretreated with a nutrient medium consisting of 50% Bacto agar 1% (Difco™, Fisher, Illkirch-Graffenstaden, France) in Gey's solution [15], 38.5% DMEM (Gibco™, Invitrogen, Paris, France), 10% fetal calf serum (Gibco™, Invitrogen), 1% L-glutamine (Gibco™, Invitrogen), and 0.05% penicillin/streptomycin solution (Gibco™, Invitrogen). Skin was isolated from 7-day-old white leghorn chicken eggs and placed in sterile PBS. The samples were cut into 1-mm<sup>2</sup> pieces and layered onto the bottom of a 70-cm<sup>2</sup> petri dish (Dominique Dutscher, Issy-les-Moulineaux, France). Nine fragments were placed in each petri dish and a 2-cm<sup>2</sup> piece of material was deposited on each of those fragments.

Each dish was used for only 1 material, and 4 dishes were prepared for each material. The cultures were incubated at 37°C with 5% CO<sub>2</sub> for 7 days, after which the materials were



**Figure 1.** (A) Organotypic culture description. (B) Method of measurement of cell migration and adhesion.

Thx, Thermanox®.

removed and stained with neutral red. Next, the total surface area covered by tissue was measured using a stereomicroscope equipped with a camera and Image J software (National Institutes of Health, Bethesda, MD, USA) [16]. This area corresponded to the total surface of the cell layer minus the initial explant surface.

Additionally, the cells were detached from the materials using 0.025% trypsin-EDTA (Gibco™, Invitrogen) in Isoton® II electrolyte solution (Beckman Coulter, Villepinte, France).

The rate of cell detachment was determined by counting the detached cells with a Multisizer® (Beckman Coulter) after 5, 10, 20, 30, and 60 minutes. The rate of detachment as a function of time was used as a measure of adhesion strength, and we defined an arbitrary index (Figure 1B).

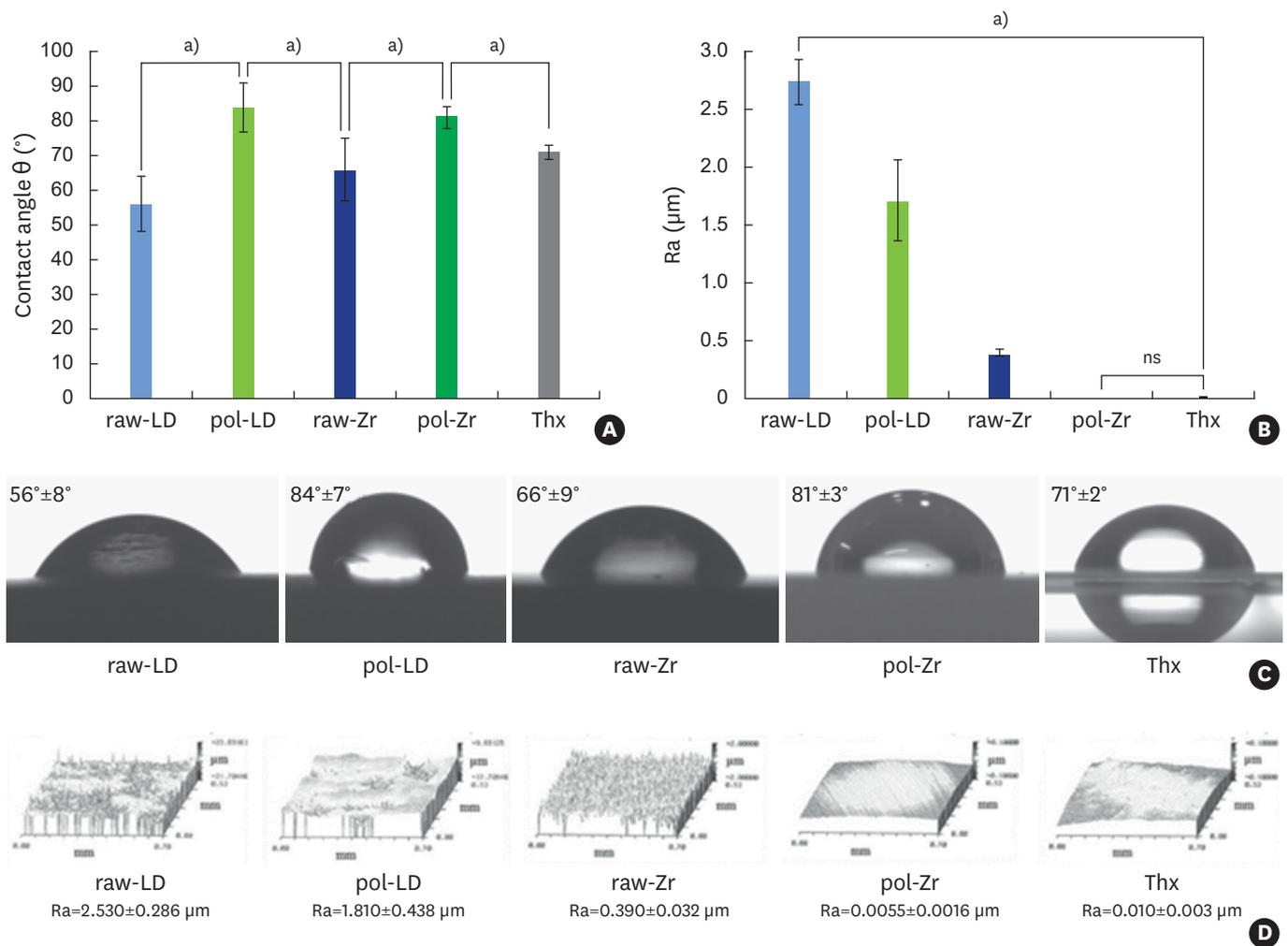
### Statistical analysis

All data were represented as means±standard error of the mean. The data were compared using 1-way analysis of variance followed by the Tukey-Kramer multiple comparisons test (InStat, GraphPad software, La Jolla, CA, USA). *P* values <0.05 were considered to indicate statistical significance.

## RESULTS

The contact angle measurements showed significantly different results between the controls and the ceramics (Figure 2A and 2C). The polishing treatment of the ceramics reduced their hydrophilicity, as illustrated by the contact angles of the water droplets, which increased from 56° to 84° for LS<sub>2</sub> ceramic sample (LD) and from 66° to 81° for ZrO<sub>2</sub> ceramic sample (Zr) following the treatment. This property is expected to influence protein adsorption and cell attachment.

The roughness of the ceramics compared to controls, as measured by scanning white-light interferometry, showed significantly different values between Thx and ceramics and between LD and Zr (Figure 2B and 2D). Pol-Zr had the same roughness as the control Thx. These results were correlated with the cell adhesion assessment. The decreasing values of roughness in raw-LD, pol-LD, raw-Zr, and pol-Zr were correlated with reduced cell adhesion.



**Figure 2.** Physicochemical analyses of raw-LD, pol-LD, raw-Zr, and pol-Zr compared to Thx. (A) Contact angle  $\theta$  (°); (B) Ra ( $\mu\text{m}$ ); (C) images of water droplets; (D) interferometry microscopy images. Error bars indicate significant differences.

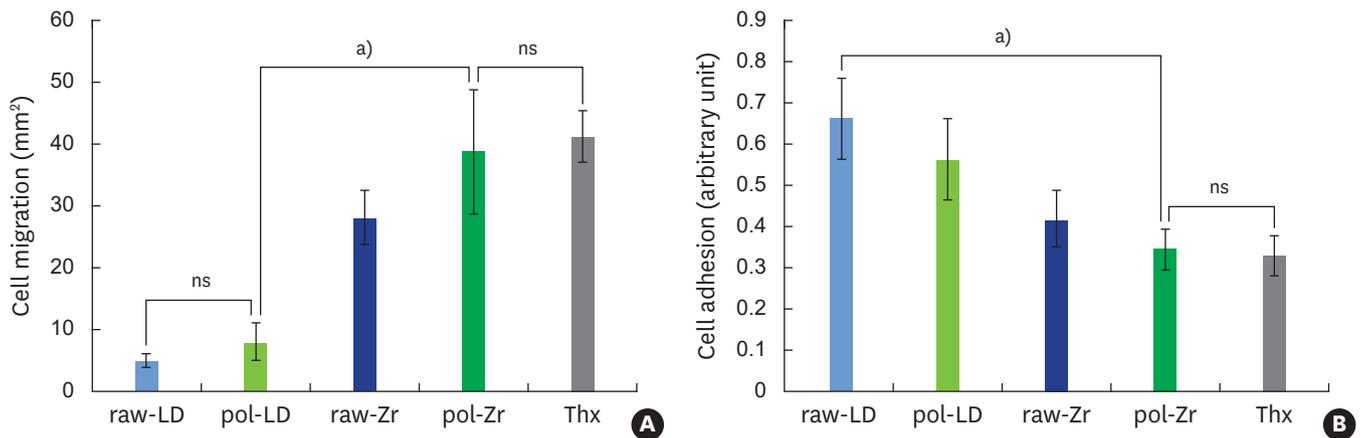
raw-Zr, raw  $\text{ZrO}_2$  ceramic sample; pol-Zr, polished  $\text{ZrO}_2$  ceramic sample; raw-LD, raw  $\text{LS}_2$  ceramic sample; pol-LD, polished  $\text{LS}_2$  ceramic sample; Thx, Thermanox®; ns, not significant; Ra, roughness.

<sup>a)</sup>Statistically significant differences ( $P < 0.001$ ).

After 7 days in organotypic culture, the epithelial layers grown on the materials (Figure 3A) showed significant differences between the control and the ceramics, and between the ceramics themselves. LD did not enhance cell migration compared to Thx, while Zr significantly increased cell migration compared to LD.

Interestingly, the cell adhesion assessment (Figure 3B) showed strong cell adhesion on the raw-LD but very weak cell adhesion on the pol-Zr and Thx ( $P < 0.001$ ). Zr (both polished and raw) showed significantly weaker cell adhesion than LD.

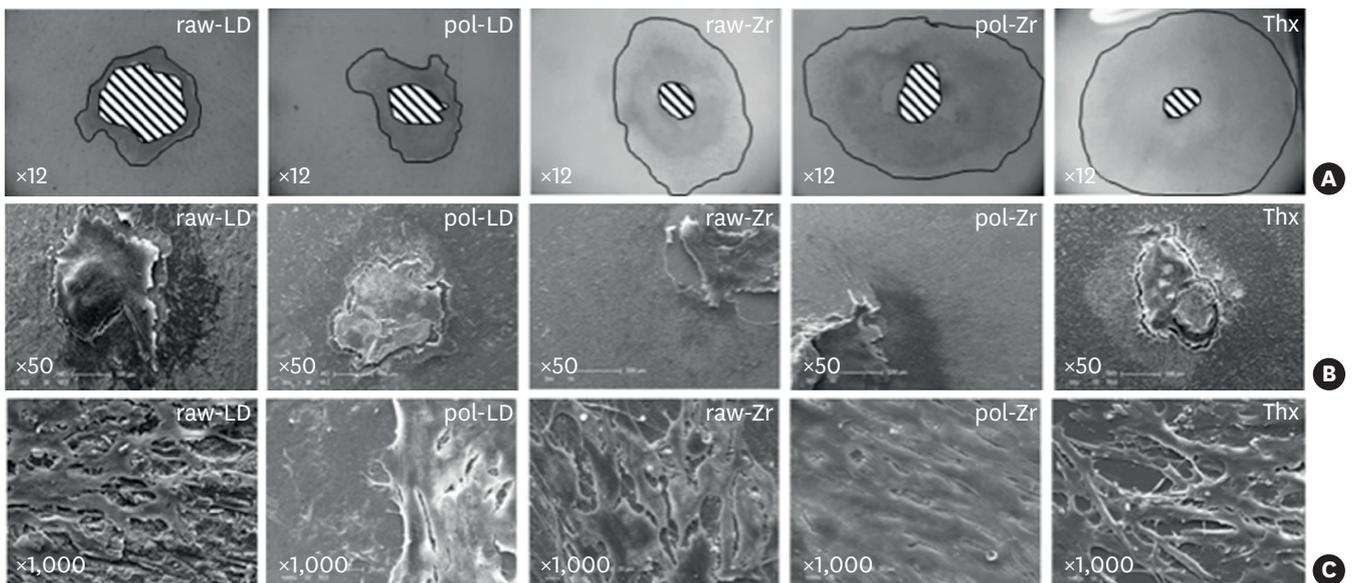
Samples observed by scanning electron microscopy (Figure 4) showed drastic differences in cell shape on the ceramics compared to Thx. A proportional increase of the cell layer area was observed in the raw-LD compared to Thx. On Thx and polished ceramics, which favored cell migration, the cells were elongated. Cells on the raw ceramics were cuboidal in shape, forming a squamous epithelium.



**Figure 3.** Organotypic culture on raw-LD, pol-LD, raw-Zr, and pol-Zr compared to Thx. (A) Cell migration (mm<sup>2</sup>); (B) cell adhesion (arbitrary unit). Error bars indicate significant differences.

raw-Zr, raw ZrO<sub>2</sub> ceramic sample; pol-Zr, polished ZrO<sub>2</sub> ceramic sample; raw-LD, raw LS<sub>2</sub> ceramic sample; pol-LD, polished LS<sub>2</sub> ceramic sample; Thx, Thermanox®; ns, not significant.

<sup>a</sup>Statistically significant differences ( $P < 0.001$ ).



**Figure 4.** Scanning results of raw-LD, pol-LD, raw-Zr, and pol-Zr compared to Thx. (A) White-light interferometry measuring the area with neutral red staining using imaging software ( $\times 12$ ); and (B, C) scanning electron microscopy showing shapes and layers of cells (B,  $\times 50$ ; C,  $\times 1,000$ ).

raw-Zr, raw ZrO<sub>2</sub> ceramic sample; pol-Zr, polished ZrO<sub>2</sub> ceramic sample; raw-LD, raw LS<sub>2</sub> ceramic sample; pol-LD, polished LS<sub>2</sub> ceramic sample; Thx, Thermanox®.

## DISCUSSION

Biocompatibility and chemical durability are highly important properties in dental materials. Zirconia ceramics have been reported not to have potential toxic or genotoxic effects [17-19] and to present satisfactory soft tissue responses [20]. Esthetic implant abutments must minimize or reduce plaque adhesion and possess light dynamic qualities comparable to natural teeth. The first objective is to allow healing around the crown and the gingiva, which requires significant roughness of the dental crown, leading to strong cell adhesion. The second objective is to avoid biofilm formation on the dental prosthesis, which involves creating surfaces that prevent both cell adhesion and protein adsorption. Dental ceramics

must offer maximal mechanical stability, outstanding biocompatibility, and accelerated and dense peri-implant soft tissue attachment [21]. They are generally required to be low-adhesive materials to avoid biofilm formation on the teeth [22].

In accordance with the periodontal environment, our organotypic culture method was used in this study instead of classic cell culture because it approaches the complexity of living tissue. With this technique, the organ in contact with the biomaterial develops a tissue comparable to that of the human gingival epithelium [23].

It is well known that migration and adhesion are biological parameters that are not necessarily directly linked. Cells can migrate rapidly with very low adhesion, which is the case for the epithelial tongue during the new-epithelialization phase of wound healing. Our approach allowed us to study the behavior of cells in direct contact with biomaterials according to these 2 parameters [23,26].

It is now widely known that cell adhesion depends on the physicochemical properties of a biomaterial through protein adsorption and extracellular matrix constitution. First, the wettability properties, assessed by the contact angle measurements, influence protein adsorption onto a biomaterial [24]. A hydrophobic surface generally induces protein adsorption and then cell adhesion, while a hydrophilic surface often prevents any adsorption and subsequent cell adhesion. Second, the roughness of a biomaterial also drastically influences protein adsorption and the behavior of the cells [25]. In this study, the cell adhesion on the different ceramics was controlled by their roughness. The biological parameters assessed using our culture model have already been demonstrated to have relationships with cell migration, cell adhesion, and physicochemical properties in other biomaterials [26].

Finally, we confirmed *in vitro* that the LD dental ceramic (in this case IPS e.max® Press, Ivoclar Vivadent SAS) was not cytotoxic [27,28] and is a promising material for improving the esthetic outcomes of dental implants and for tightening the peri-implant junction [23] due to simple surface modifications making it possible to create 2 different types of interfaces on a single material. The abrasion provides micro-rough surfaces with strong wettability, which allows strong adhesion of epithelial tissues. This type of surface will be optimal for gingival adhesion around the dental abutment. In contrast, applying the glazing technique to the same material will provide a smooth surface with strong hydrophilicity [23]; which gives restricted adhesion properties to the material, as appropriate for a dental surface designed to prevent biofilm formation in the septic environment of the mouth [29].

The key to an esthetically pleasing appearance lies in the clinician's ability to properly manage the soft tissue profile around dental implants [30]. An all-ceramic system with LS<sub>2</sub> ceramic could improve the final esthetic results. Our study clearly demonstrates that it is easily possible, by simple surface treatments that can be performed by a practitioner, to modify the properties of the ceramics used in dentistry. These results demonstrate how simple surface modifications (i.e., polished vs. raw) can finely modulate tissue adhesion.

Dental surgeons need to choose the best surface treatment for each clinical application, especially in dental implantology. Soft tissue management for the visible dental zone could be improved by choosing LD press ceramics as an esthetic and biocompatible material for a tailor-made, manually modifiable abutment.

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